



## **Smoothelin, a new marker to determine the origin of liver fibrogenic cells**

Lepreux, Sébastien ; Guyot, Christelle ; Billet, Fabrice ; Combe, Chantal ; Balabaud, Charles ;  
Bioulac-Sage, Paulette ; Desmoulière, Alexis

**Abstract:** AIM: To explore this hypothesis that smooth muscle cells may be capable of acquiring a myofibroblastic phenotype, we have studied the expression of smoothelin in fibrotic conditions. **METHODS:** Normal liver tissue (n = 3) was obtained from macroscopically normal parts of hepatectomy, taken at a distance from hemangiomas. Pathological specimens included post-burn cutaneous hypertrophic scars (n = 3), fibrotic liver tissue (n = 5), cirrhotic tissue (viral and alcoholic hepatitis) (n = 5), and hepatocellular carcinomas (n = 5). Tissue samples were fixed in 10% formalin and embedded in paraffin for immunohistochemistry or were immediately frozen in liquid nitrogen-cooled isopentane for confocal microscopy analysis. Sections were stained with antibodies against smoothelin, which is expressed exclusively by smooth muscle cells, and  $\alpha$ -smooth muscle actin, which is expressed by both smooth muscle cells and myofibroblasts. **RESULTS:** In hypertrophic scars,  $\alpha$ -smooth muscle actin was detected in vascular smooth muscle cells and in numerous myofibroblasts present in and around nodules, whereas smoothelin was exclusively expressed in vascular smooth muscle cells. In the normal liver, vascular smooth muscle cells were the only cells that express  $\alpha$ -smooth muscle actin and smoothelin. In fibrotic areas of the liver, myofibroblasts expressing  $\alpha$ -smooth muscle actin were detected. Myofibroblasts co-expressing  $\alpha$ -smooth muscle actin and smoothelin were observed, and their number was slightly increased in parallel with the degree of fibrosis (absent in liver with mild or moderate fibrosis; 5% to 10% positive in liver showing severe fibrosis). In cirrhotic septa, numerous myofibroblasts co-expressed  $\alpha$ -smooth muscle actin and smoothelin (more than 50%). In hepatocellular carcinomas, the same pattern of expression for  $\alpha$ -smooth muscle actin and smoothelin was observed in the stroma reaction surrounding the tumor and around tumoral cell plates. In all pathological liver samples,  $\alpha$ -smooth muscle actin and smoothelin were co-expressed in vascular smooth muscle cells. **CONCLUSION:** During development of advanced liver fibrosis, a sub-population of myofibroblasts expressing smoothelin may be derived from vascular smooth muscle cells, illustrating the different cellular origins of myofibroblasts.

DOI: <https://doi.org/10.3748/wjg.v19.i48.9343>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-93488>

Journal Article

Published Version

Originally published at:

Lepreux, Sébastien; Guyot, Christelle; Billet, Fabrice; Combe, Chantal; Balabaud, Charles; Bioulac-Sage, Paulette; Desmoulière, Alexis (2013). Smoothelin, a new marker to determine the origin of liver fibrogenic cells. *World Journal of Gastroenterology*, 19(48):9343-9350.

DOI: <https://doi.org/10.3748/wjg.v19.i48.9343>



## Smoothelin, a new marker to determine the origin of liver fibrogenic cells

Sébastien Lepreux, Christelle Guyot, Fabrice Billet, Chantal Combe, Charles Balabaud, Paulette Bioulac-Sage, Alexis Desmoulière

Sébastien Lepreux, CHU Bordeaux, Hôpital Pellegrin, Service d'Anatomie Pathologique, F-33076 Bordeaux, France  
Christelle Guyot, Chantal Combe, Charles Balabaud, Paulette Bioulac-Sage, U889 and Université Victor Segalen Bordeaux 2, F-33076 Bordeaux, France

Christelle Guyot, Division of Clinical Pharmacology and Toxicology, Department of Medicine, University Hospital Zurich, 8006 Zurich, Switzerland

Fabrice Billet, Alexis Desmoulière, Facultés de Médecine et de Pharmacie, Université de Limoges, EA 6309, F-87025 Limoges, France

**Author contributions:** Lepreux S and Guyot C contributed equally to this work; Lepreux S, Guyot C and Combe C performed the research; Billet F, Balabaud C and Bioulac-Sage P contributed to new reagents/analytic tools and analyzed the data; Lepreux S, Guyot C and Desmoulière A designed the research and wrote the paper.

**Supported by** In part a grant from the French Ministry of Research

**Correspondence to:** Alexis Desmoulière, PhD, Faculté de Pharmacie, Université de Limoges, 2 rue du Docteur Marcland, EA 6309, F-87025 Limoges, France. [alexis.desmouliere@unilim.fr](mailto:alexis.desmouliere@unilim.fr)

Telephone: +33-555-435873 Fax: +33-555-435912

Received: May 3, 2013 Revised: June 18, 2013

Accepted: July 4, 2013

Published online: December 28, 2013

### Abstract

**AIM:** To explore this hypothesis that smooth muscle cells may be capable of acquiring a myofibroblastic phenotype, we have studied the expression of smoothelin in fibrotic conditions.

**METHODS:** Normal liver tissue ( $n = 3$ ) was obtained from macroscopically normal parts of hepatectomy, taken at a distance from hemangiomas. Pathological specimens included post-burn cutaneous hypertrophic scars ( $n = 3$ ), fibrotic liver tissue ( $n = 5$ ), cirrhotic tissue (viral and alcoholic hepatitis) ( $n = 5$ ), and hepatocellular carcinomas ( $n = 5$ ). Tissue samples were fixed

in 10% formalin and embedded in paraffin for immunohistochemistry or were immediately frozen in liquid nitrogen-cooled isopentane for confocal microscopy analysis. Sections were stained with antibodies against smoothelin, which is expressed exclusively by smooth muscle cells, and  $\alpha$ -smooth muscle actin, which is expressed by both smooth muscle cells and myofibroblasts.

**RESULTS:** In hypertrophic scars,  $\alpha$ -smooth muscle actin was detected in vascular smooth muscle cells and in numerous myofibroblasts present in and around nodules, whereas smoothelin was exclusively expressed in vascular smooth muscle cells. In the normal liver, vascular smooth muscle cells were the only cells that express  $\alpha$ -smooth muscle actin and smoothelin. In fibrotic areas of the liver, myofibroblasts expressing  $\alpha$ -smooth muscle actin were detected. Myofibroblasts co-expressing  $\alpha$ -smooth muscle actin and smoothelin were observed, and their number was slightly increased in parallel with the degree of fibrosis (absent in liver with mild or moderate fibrosis; 5% to 10% positive in liver showing severe fibrosis). In cirrhotic septa, numerous myofibroblasts co-expressed  $\alpha$ -smooth muscle actin and smoothelin (more than 50%). In hepatocellular carcinomas, the same pattern of expression for  $\alpha$ -smooth muscle actin and smoothelin was observed in the stroma reaction surrounding the tumor and around tumoral cell plates. In all pathological liver samples,  $\alpha$ -smooth muscle actin and smoothelin were co-expressed in vascular smooth muscle cells.

**CONCLUSION:** During development of advanced liver fibrosis, a subpopulation of myofibroblasts expressing smoothelin may be derived from vascular smooth muscle cells, illustrating the different cellular origins of myofibroblasts.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

**Key words:** Smooth muscle cells; Myofibroblasts;  $\alpha$ -smooth muscle actin; Smoothelin; Fibrosis/cirrhosis; Hepatocellular carcinoma

**Core tip:** In fibrotic conditions, it has been suggested that smooth muscle cells can acquire a myofibroblastic phenotype. To explore this hypothesis, we studied the expression of smoothelin, a specific marker of end-stage differentiation of smooth muscle cells, in cutaneous and hepatic fibrotic conditions, using immunohistochemistry and confocal microscopy. We showed that during advanced liver fibrosis, a subpopulation of  $\alpha$ -smooth muscle actin-expressing myofibroblasts also express smoothelin and thus may be derived from vascular smooth muscle cells. This finding, which illustrates the different potential cellular origins of myofibroblasts involved in liver fibrogenesis, may represent an interesting tool to distinguish advanced stages of cirrhosis.

Lepreux S, Guyot C, Billet F, Combe C, Balabaud C, Bioulac-Sage P, Desmoulière A. Smoothelin, a new marker to determine the origin of liver fibrogenic cells. *World J Gastroenterol* 2013; 19(48): 9343-9350 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v19/i48/9343.htm> DOI: <http://dx.doi.org/10.3748/wjg.v19.i48.9343>

## INTRODUCTION

Smoothelin, a constituent of the smooth muscle cell cytoskeleton, has been described as a marker of end-stage differentiation of smooth muscle cells because it has been found only in contractile smooth muscle cells<sup>[1-3]</sup>. Smoothelin has two major isoforms in adults, which are expressed in a tissue specific manner: a 59 kDa isoform, smoothelin-A, which is expressed in visceral and urogenital tissues, such as the digestive tract, bladder, and prostate; and a 110 kDa isoform, smoothelin-B, which is expressed in blood vessel walls<sup>[4]</sup>. Transient synthesis of a third smoothelin isoform has been detected in embryonic striated muscle cells in chicken<sup>[5]</sup>. In cultured smooth muscle cells, smoothelin colocalizes with  $\alpha$ -smooth muscle actin stress fibers<sup>[3]</sup> and smoothelin can bind to  $\alpha$ -smooth muscle actin, which suggested a direct role of smoothelin in contraction<sup>[6]</sup>. In contrast, cells with smooth muscle cell-like features, such as myofibroblasts, do not express smoothelin<sup>[3]</sup>. Myofibroblasts are contractile cells that express  $\alpha$ -smooth muscle actin<sup>[7]</sup>. These cells are involved in tissue repair processes and, particularly, in extracellular matrix deposition and remodeling<sup>[8]</sup>. In normal connective tissues, myofibroblasts are rare<sup>[9]</sup>. After tissue injury, myofibroblasts appear, and it is commonly accepted that most are derived from locally recruited connective tissue fibroblasts<sup>[10]</sup>. In the liver, the most important cells involved in fibrogenesis are hepatic stellate cells and portal fibroblasts, which are able to differentiate into myofibroblasts and are responsible for matrix deposition<sup>[11]</sup>. However, the involvement of different cell

types with various origins, such as smooth muscle cells, circulating cells or bone marrow-derived cells has also been suggested in the establishment of liver fibrosis/cirrhosis<sup>[12-16]</sup>. However, the degree to which this process contributes to fibrosis remains a matter of intense debate and is likely to be context-dependent. To determine the possible contribution of smooth muscle cells to the appearance of myofibroblasts during fibrotic processes, we studied the expression of smoothelin in different diseases that show myofibroblast involvement; *i.e.*, post-burn cutaneous hypertrophic scars, liver fibrosis and cirrhosis, and hepatocellular carcinomas.

## MATERIALS AND METHODS

### Human liver samples and tissue processing

Human tissue samples used in this study were selected from the files of the tissue bank of the Department of Pathology (CHU Bordeaux, Pellegrin Hospital, Bordeaux, France). Normal liver tissues ( $n = 3$ ) were obtained from macroscopically normal parts of livers after hepatectomy, taken at a distance from hemangiomas. Pathological specimens included post-burn cutaneous hypertrophic scars ( $n = 3$ ); fibrotic liver tissue ( $n = 5$ ), ranging from F1 to F3 stage according to the Metavir score<sup>[17]</sup>; cirrhotic tissues (viral and alcoholic hepatitis) ( $n = 5$ ); and hepatocellular carcinomas ( $n = 5$ ). Tissue samples were fixed in 10% buffered formalin, embedded in paraffin and processed for diagnostic purposes and immunohistochemistry, or were immediately frozen in liquid nitrogen-cooled isopentane for confocal microscopy analysis. The procedures were carried out in accordance with the European Guidelines for the use of human tissues.

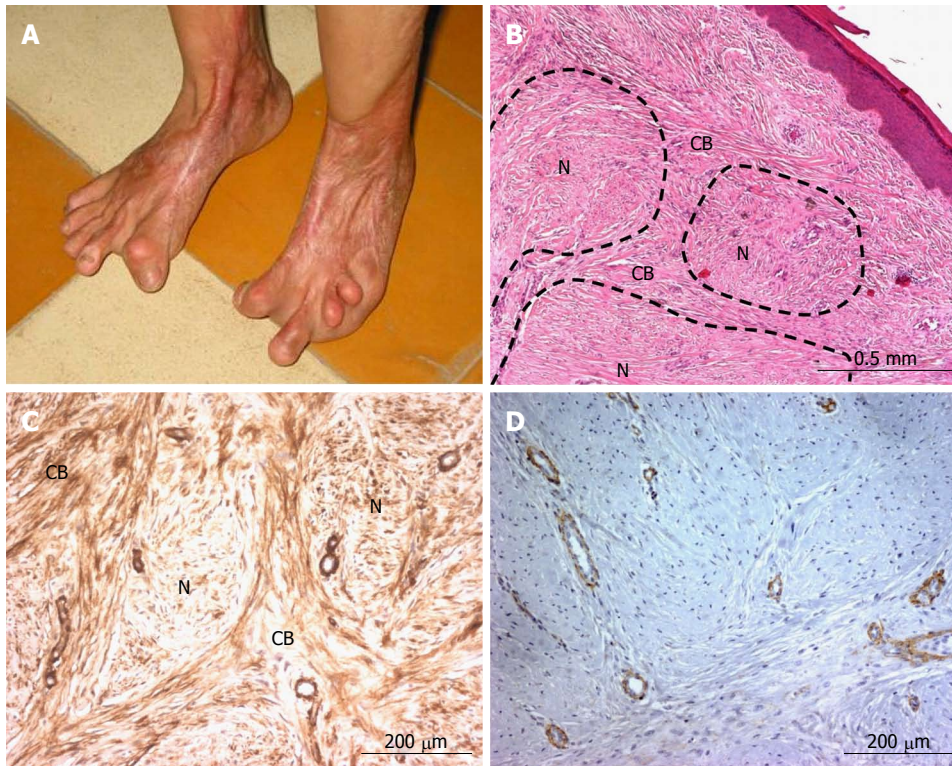
### Immunohistochemistry and confocal microscopy

Mouse monoclonal antibodies against smoothelin (immunoglobulin G, IgG1 clone R4A, which reacts with smoothelin A and B, MUBio Products, Maastricht, The Netherlands), and  $\alpha$ -smooth muscle actin (IgG2a clone 1A4, Dako SA, Trappes, France) were used. These two antibodies have been extensively used and their specificity has been clearly documented<sup>[1,18]</sup>. Immunohistochemistry was essentially performed as previously described<sup>[19]</sup>. Briefly, after incubation with the first antibody, the epitopes were detected using the Vectastain<sup>®</sup> ABC system (Vector Laboratories, Peterborough, United Kingdom) or the Envision<sup>™</sup> system (DakoCytomation, Trappes, France), with diaminobenzidine as the color substrate. Slides were then counterstained with hematoxylin.

For double immunofluorescence, cryostat sections were first incubated with the antibodies against smoothelin and  $\alpha$ -smooth muscle actin, then with a TRITC-conjugated goat anti-mouse IgG1 (Southern Biotech, Birmingham, AL, United States) and an Alexa Fluor<sup>®</sup> 488 goat anti-mouse IgG2a (Molecular Probes, Eugene, OR, United States). Nuclei were stained with 4',6-diamidino-2-phenylindole (DAPI).

The specificity of staining was confirmed by incubation





**Figure 1**  $\alpha$ -smooth muscle actin and smoothelin expression in a cutaneous hypertrophic scar. A: Following a burn injury, a retractor hypertrophic scar was observed (from Vincent Casoli, Plastic Surgery and Burns Unit, University Hospital of Bordeaux, France); B: Hematoxylin and eosin staining shows typical hypertrophic scar architecture with nodules (N) surrounded by cell bundles (CB); C:  $\alpha$ -smooth muscle actin is expressed in vascular smooth muscle cells and myofibroblasts both in and around the nodules; D: Smoothelin is only expressed by vascular tunica media smooth muscle cells.

tion in non-immune serum and in the absence of the primary antibody. For immunohistochemistry, sections were examined with a Zeiss Axioplan 2 microscope (Carl Zeiss Microscopy, Jena, Germany). Images were acquired with an AxioCam camera (Carl Zeiss Vision, Hallbergmoos, Germany) by means of the AxioVision image processing and analysis system (Carl Zeiss Vision). For double immunofluorescence, sections were analyzed with a confocal microscopy (LSM Meta 510, Carl Zeiss Microscopy).

For quantitative evaluation of staining, cell counting was performed in the septa of fibrotic and cirrhotic livers, and in the stroma reaction of hepatocellular carcinoma. Vessels were not included in this evaluation. For each field, the ratio of the number of smoothelin-positive and  $\alpha$ -smooth muscle actin-positive cells over the number of cells only expressing  $\alpha$ -smooth muscle actin was calculated. The analysis was performed on an average of 10 fields/zone using the  $\times 40$  objective. Only positive cells containing a nucleus were counted.

## RESULTS

### $\alpha$ -smooth muscle actin and smoothelin expression in hypertrophic scars

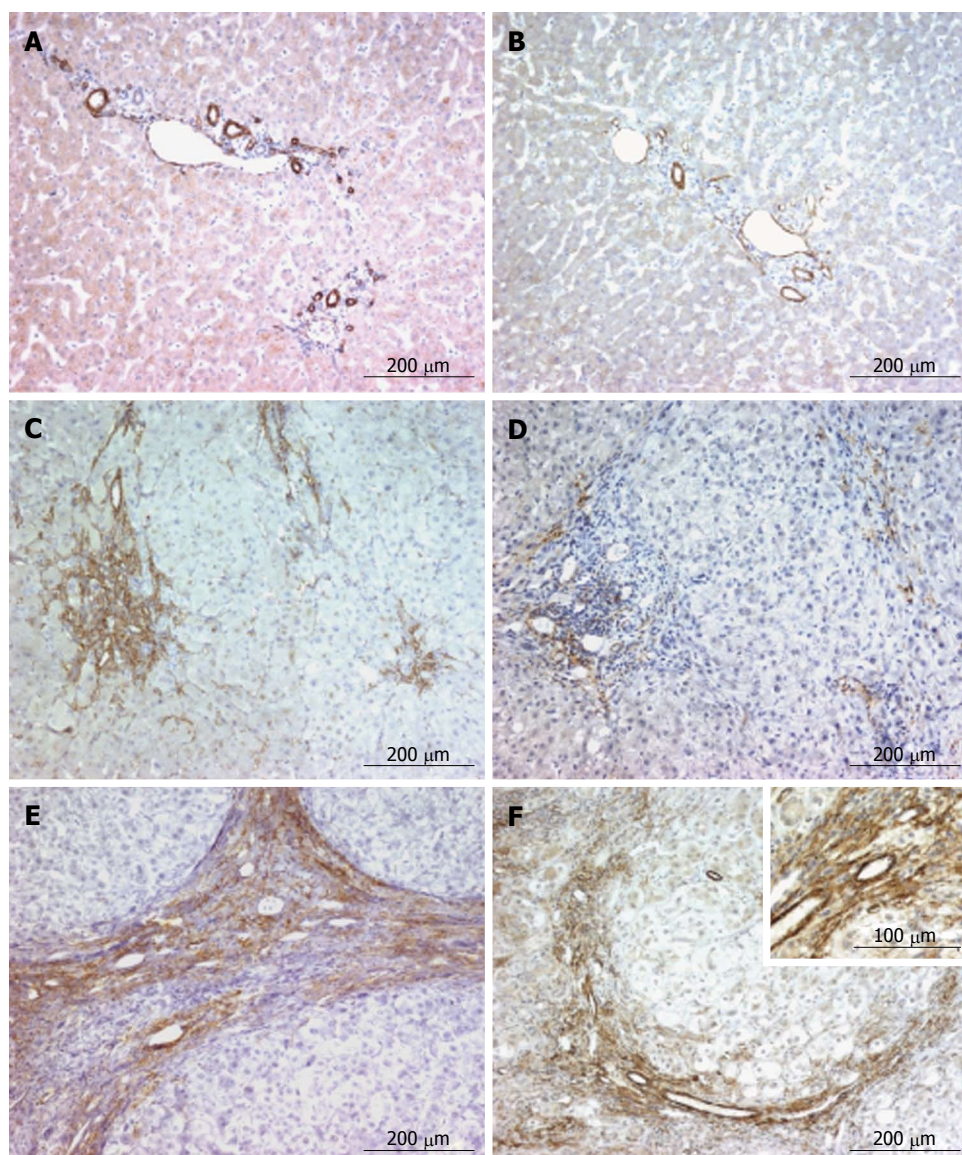
In many situations, hypertrophic scars develop significant contractile activity (because of the presence of myofibroblasts) (Figure 1A), and are usually organized in a nodular pattern (Figure 1B). By immunohistochemistry,  $\alpha$ -smooth

muscle actin was detected in blood vessel walls and in numerous myofibroblasts present in and around the nodules (Figure 1C), whereas smoothelin was exclusively shown in the vessel walls (Figure 1D). Our data showed that myofibroblasts present in hypertrophic scars do not express smoothelin.

### $\alpha$ -smooth muscle actin and smoothelin expression in normal and pathological livers

In the normal liver,  $\alpha$ -smooth muscle actin was exclusively expressed by the smooth muscle cells within the tunica media of portal arteries and veins, and of the centrilobular veins (Figure 2A). The expression of smoothelin in the normal liver was also detected in the smooth muscle cells of these vessels (Figure 2B). Smoothelin and  $\alpha$ -smooth muscle actin showed a similar distribution.

In livers with mild or moderate fibrosis (F1-F2), myofibroblasts present in fibrotic areas clearly expressed  $\alpha$ -smooth muscle actin, but did not express smoothelin (data not shown). In livers showing severe fibrosis (F3), strong staining for  $\alpha$ -smooth muscle actin was detected in myofibroblasts present in fibrotic areas (Figure 2C). Weak expression of smoothelin was present in a few myofibroblasts (Figure 2D). In cirrhotic livers (F4), myofibroblasts present in cirrhotic septa expressed high amounts of  $\alpha$ -smooth muscle actin (Figure 2E). Numerous myofibroblasts also expressed smoothelin (Figure 2F). In all stages of liver fibrosis,  $\alpha$ -smooth muscle actin



**Figure 2**  $\alpha$ -smooth muscle actin and smoothelin expression in normal, fibrotic and cirrhotic livers. A and B: In the normal liver,  $\alpha$ -smooth muscle actin (A) and smoothelin (B) show similar expression in tunica media smooth muscle cells of the portal arteries and veins and in the centrilobular veins; C and D: In severe fibrosis, myofibroblasts express high amounts of  $\alpha$ -smooth muscle actin (C), while smoothelin is only expressed at low levels (D) ( $\alpha$ -smooth muscle actin and smoothelin are coexpressed in vascular smooth muscle cells); E and F: In the cirrhotic liver, myofibroblasts express  $\alpha$ -smooth muscle actin in fibrotic septae (E) and numerous myofibroblasts also express smoothelin (F); however, smoothelin expression is lower compared with  $\alpha$ -smooth muscle actin expression (insert).  $\alpha$ -smooth muscle actin and smoothelin are co-expressed in vascular tunica media smooth muscle cells (F).

and smoothelin were co-expressed in vascular smooth muscle cells (Figure 2C-F).

The co-localization of  $\alpha$ -smooth muscle actin and of smoothelin in the same cells was confirmed by double immunofluorescence (Figure 3). All cells expressing smoothelin also expressed  $\alpha$ -smooth muscle actin, including vascular smooth muscle cells and myofibroblasts. Moreover, a quantitative evaluation revealed that in livers showing severe fibrosis (F3), 5% to 10% of myofibroblasts co-expressed  $\alpha$ -smooth muscle actin and smoothelin. In cirrhotic septa, more than 50% of myofibroblasts co-expressed  $\alpha$ -smooth muscle actin and smoothelin.

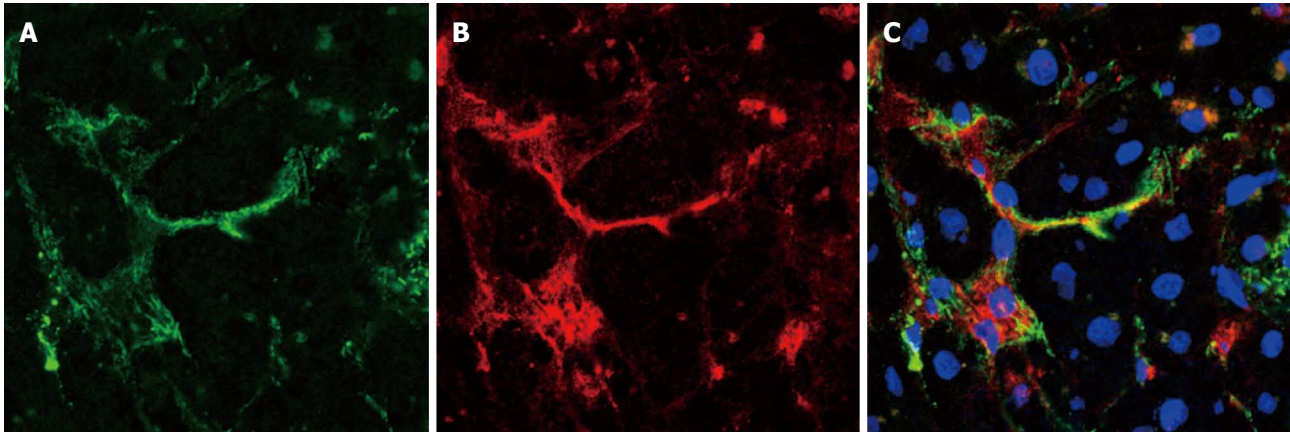
In hepatocellular carcinomas, we observed the same expression pattern of  $\alpha$ -smooth muscle actin and smoothelin in the stroma reaction surrounding the tumor (Figure

4A and B). These two proteins were also expressed by fusiform pericyte-like cells between endothelial cells of the sinusoidal capillaries and tumoral cell plates (Figure 4C and D).

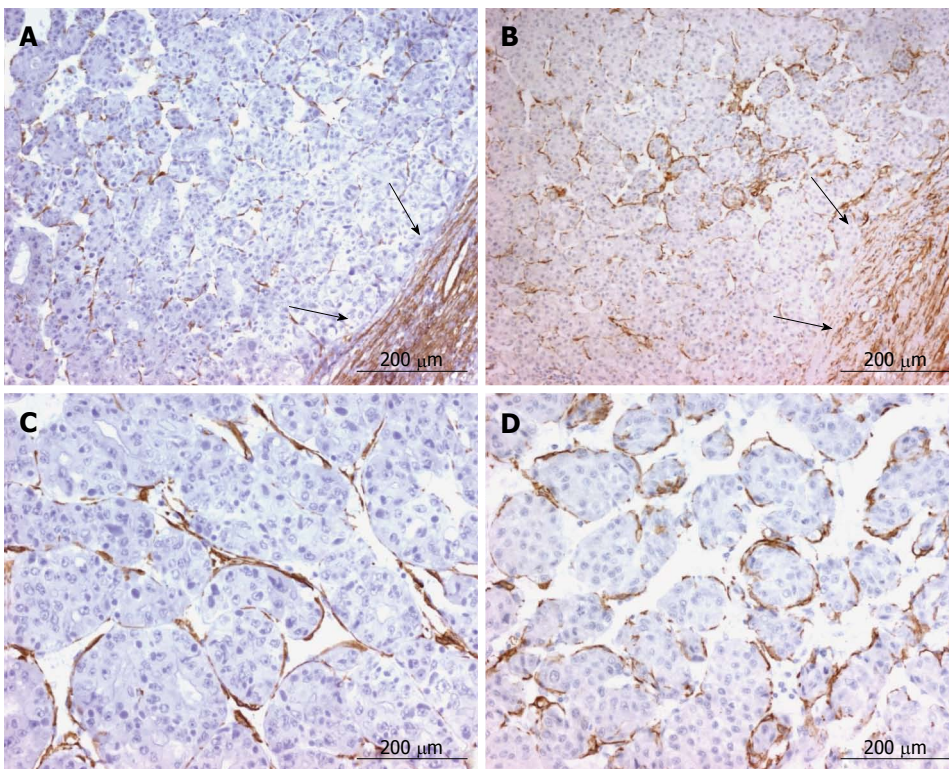
## DISCUSSION

In normal conditions, several cell types that have contractile properties express  $\alpha$ -smooth muscle actin, such as smooth muscle cells and, to a lesser extent, pericytes. Smooth muscle cells display a large variation in phenotype among, and even within, tissues and organs. Different patterns of marker expression reflect, in part, the heterogeneity of smooth muscle cell subpopulations and their phenotypic modulation<sup>[20]</sup>. Smoothelin, a recently





**Figure 3** Cellular co-localization of  $\alpha$ -smooth muscle actin and smoothelin in a fibrotic liver. A:  $\alpha$ -smooth muscle actin expression (green); B: Smoothelin expression (red); C: Merged (nuclei are stained with DAPI). Smoothelin-expressing cells also express  $\alpha$ -smooth muscle actin.



**Figure 4**  $\alpha$ -Smooth muscle actin and smoothelin expression in hepatocellular carcinoma. A and B:  $\alpha$ -smooth muscle actin (A) and smoothelin (B) are expressed similarly in the stroma reaction surrounding the tumor (arrows) and around tumoral hepatocytes; C and D:  $\alpha$ -smooth muscle actin (C) and smoothelin (D) are expressed by pericyte-like cells underlying capillaries between tumoral hepatocytes.

described smooth muscle cell marker, illustrates these phenotypic features. Firstly, smoothelin expression varies, with isoform A being expressed in visceral smooth muscle cells, and isoform B being expressed in vascular smooth muscle cells<sup>[4]</sup>. Secondly, smoothelin is expressed only by fully differentiated smooth muscle cells and not by proliferative or non-contractile smooth muscle cells<sup>[1]</sup>. Thirdly, visceral smoothelin expression is different according to the location of the smooth muscle cells within the organ. For example, in the digestive tract and urinary tract, most of the smooth muscle cells of the muscularis propria express smoothelin, but the smooth muscle cells

of the muscularis mucosae do not<sup>[21,22]</sup>. During fibrotic processes, it is assumed that it is mainly fibroblasts that are recruited and acquire a myofibroblastic phenotype, *i.e.*, a smooth muscle cell-like phenotype. However, it has been suggested that smooth muscle cells can also contribute to the appearance of myofibroblasts. Similarly to smooth muscle cells, fully differentiated myofibroblasts express  $\alpha$ -smooth muscle actin and are contractile cells; in addition, myofibroblasts are responsible for extracellular matrix deposition. Like myofibroblasts observed within bladder carcinoma<sup>[21]</sup>, blood vessel adventitia<sup>[23]</sup> or the airway wall of patients with asthma<sup>[24]</sup>, our study shows

that myofibroblasts in cutaneous hypertrophic scars do not express smoothelin. In the skin, myofibroblasts are mostly derived from locally recruited connective tissue fibroblasts, which, prior to injury, do not express  $\alpha$ -smooth muscle actin or smoothelin<sup>[8]</sup>. These fibroblasts, when transformed into myofibroblasts, express  $\alpha$ -smooth muscle actin but not smoothelin. Additionally, it has been shown that, in hypertrophic scars, a population of fibrogenic circulating cells, called fibrocytes, is recruited and participates in scar formation<sup>[25]</sup>. Here, we showed that, in hypertrophic scars, myofibroblasts do not express smoothelin, which suggests that these myofibroblasts probably do not derive from local smooth muscle cells. However, our study showed that, in fibrotic liver diseases, myofibroblasts can express smoothelin. Double immunofluorescence and confocal analysis confirmed the co-expression of  $\alpha$ -smooth muscle actin and smoothelin in the same cells. This expression is stronger in parallel with the progression and the chronicity of the fibrotic lesion: i.e., there are more smoothelin-expressing myofibroblasts within advanced stages of fibrosis or cirrhosis than in the first stages of fibrosis. Smoothelin expression may thus appear gradually during fibrosis development; therefore, smoothelin could represent a good marker for evaluating the prognosis of liver fibrosis. In the liver, numerous cell types can be recruited and acquire a myofibroblastic phenotype; generally these are fibroblasts within the portal tract and hepatic stellate cells in the space of Disse<sup>[11]</sup>. As shown here, these cells do not express smoothelin. However, myofibroblasts can also originate from the smooth muscle cells present in portal vessels and centrilobular veins<sup>[11]</sup>, which, as described here, do express smoothelin. In chronic hepatic schistosomiasis, it has been suggested that smooth muscle cells from the artery and vein tunica media are involved in the portal fibrogenesis that occurs in this pathology<sup>[12]</sup>. In the experimental model of porcine serum-induced fibrosis in rats, mesenchymal cells, located around the centrilobular vein, called second layer cells, participate in the formation of fibrotic septa<sup>[26]</sup>. The expression of smoothelin within smooth muscle cells can also vary. In early atherosclerotic lesions, smoothelin expression decreases in the vicinity of the affected area<sup>[27,28]</sup>; however, when the plaques become “quiescent”, smoothelin expression can be detected again<sup>[2]</sup>. Finally, we cannot exclude the possibility that the expression of smoothelin within septal myofibroblastic cells is a residual smooth muscle feature, underlining the possible smooth muscle cell origin of a subpopulation of myofibroblasts. Lastly, the tumoral stroma is the connective tissue surrounding tumoral cells<sup>[29]</sup>. No smoothelin-expressing stromal cells were described in published studies of bladder carcinoma<sup>[21,30]</sup>. In hepatocellular carcinoma, we show that numerous  $\alpha$ -smooth muscle actin expressing myofibroblasts in the stroma reaction also express smoothelin, underlining the possible smooth muscle origin of these myofibroblasts. Surprisingly, we also showed that, in hepatocellular carcinomas, cells located between endothelial cells and tumoral hepatocytes express  $\alpha$ -smooth muscle actin and smoothelin. Pericytes and hepatic stellate cells,

also termed liver-specific pericytes<sup>[31,32]</sup>, can express  $\alpha$ -smooth muscle actin, but not smoothelin<sup>[2]</sup>. This pattern of smoothelin expression, which was not found in the fibrotic liver, illustrates the complete reorganization of the capillarized liver sinusoids during tumor development, with the appearance of specific cells, and may represent an additional feature for diagnosing hepatocellular carcinomas. In summary, we hypothesize that during liver fibrosis, a subpopulation of myofibroblasts express smoothelin. These myofibroblasts may be derived from smooth muscle cells coming from vascular walls, which are recruited to participate in the fibrotic process. These cells expressing smoothelin are recruited in advanced stages of cirrhosis, when a significant vascular reorganization occurs, including portovenous and arteriovenous shunting; therefore, smoothelin could be a useful marker to distinguish advanced stages of cirrhosis. In addition, in hepatocellular carcinomas, smoothelin expression shows a very specific pattern. These data again illustrate the different cellular origins of the so-called myofibroblastic cells involved in liver fibrogenesis.

## COMMENTS

### Background

Myofibroblasts are fibrogenic and contractile cells involved in remodeling and healing processes. In most organs, myofibroblasts are rare in normal physiological conditions. After tissue injury, myofibroblasts appear, which are derived from local stromal cells, such as resident fibroblasts or organ specific stromal cells, such as hepatic stellate cells in the liver.

### Research frontiers

To explore the origin of myofibroblasts, the authors studied the expression of smoothelin, a constituent of the smooth muscle cell cytoskeleton, in normal and fibrotic livers, in hepatocellular carcinoma and in a cutaneous hypertrophic scar. Immunohistochemistry was used to identify smoothelin-expressing cells.

### Innovations and breakthroughs

This is the first study exploring smoothelin expression in the normal liver, fibrotic liver and in hepatocellular carcinoma. The results show an increase in expression that correlated with the degree of fibrosis, suggesting the progressive involvement of resident smooth muscle cells in myofibroblast recruitment. In hepatocellular carcinoma, smoothelin is expressed by pericyte-like cells between the endothelial cells of the capillaries and the tumoral cell plates.

### Applications

This preliminary study underlines that tracing the origin of myofibroblasts may be useful to evaluate the degree of fibrosis in the liver and the level of reorganization in hepatocellular carcinomas.

### Terminology

Smoothelin is a constituent of the smooth muscle cell cytoskeleton. It has been described as a marker of end-stage differentiation of smooth muscle cells because its expression has been found only in contractile smooth muscle cells.

### Peer review

This is a good descriptive study in which the authors explore the expression pattern of a smooth muscle marker, smoothelin, within normal, fibrotic and tumoral livers, as well as in a cutaneous hypertrophic scar. The results are interesting and suggest that smoothelin expression increases with the degree of fibrosis, allowing the evaluation of the transition between fibrosis and cirrhosis. Moreover, in hepatocellular carcinoma, smoothelin is expressed by some stromal cells present beneath endothelial cells of the capillary surrounding the tumoral cell plates and this distinctive feature could be useful for hepatocellular carcinoma diagnosis.

## REFERENCES

- 1 van der Loop FT, Schaart G, Timmer ED, Ramaekers FC,



- van Eys GJ. Smoothelin, a novel cytoskeletal protein specific for smooth muscle cells. *J Cell Biol* 1996; **134**: 401-411 [PMID: 8707825]
- 2 **van der Loop FT**, Gabbiani G, Kohnen G, Ramaekers FC, van Eys GJ. Differentiation of smooth muscle cells in human blood vessels as defined by smoothelin, a novel marker for the contractile phenotype. *Arterioscler Thromb Vasc Biol* 1997; **17**: 665-671 [PMID: 9108778 DOI: 10.1161/01.ATV.17.4.665]
  - 3 **Christen T**, Verin V, Bochaton-Piallat M, Popowski Y, Ramaekers F, Debruyne P, Camenzind E, van Eys G, Gabbiani G. Mechanisms of neointima formation and remodeling in the porcine coronary artery. *Circulation* 2001; **103**: 882-888 [PMID: 11171799 DOI: 10.1161/01.CIR.103.6.882]
  - 4 **van Eys GJ**, Völler MC, Timmer ED, Wehrens XH, Small JV, Schalken JA, Ramaekers FC, van der Loop FT. Smoothelin expression characteristics: development of a smooth muscle cell in vitro system and identification of a vascular variant. *Cell Struct Funct* 1997; **22**: 65-72 [PMID: 9113392 DOI: 10.1247/csf.22.65]
  - 5 **Deruiter MC**, Rensen SS, Coolen GP, Hierck BP, Bergwerff M, Debie WM, Gittenberger-De Groot AC, Van Eys GJ. Smoothelin expression during chicken embryogenesis: detection of an embryonic isoform. *Dev Dyn* 2001; **221**: 460-463 [PMID: 11500983 DOI: 10.1002/dvdy.1156]
  - 6 **Niessen P**, Rensen S, van Deursen J, De Man J, De Laet A, Vanderwinden JM, Wedel T, Baker D, Doevedans P, Hofker M, Gijbels M, van Eys G. Smoothelin-a is essential for functional intestinal smooth muscle contractility in mice. *Gastroenterology* 2005; **129**: 1592-1601 [PMID: 16285958 DOI: 10.1053/j.gastro.2005.08.018]
  - 7 **Tomasek JJ**, Gabbiani G, Hinz B, Chaponnier C, Brown RA. Myofibroblasts and mechano-regulation of connective tissue remodelling. *Nat Rev Mol Cell Biol* 2002; **3**: 349-363 [PMID: 11988769 DOI: 10.1038/nrm809]
  - 8 **Sarrazy V**, Billet F, Micallef L, Coulomb B, Desmoulière A. Mechanisms of pathological scarring: role of myofibroblasts and current developments. *Wound Repair Regen* 2011; **19** Suppl 1: s10-s15 [PMID: 21793960 DOI: 10.1111/j.1524-475X.2011.00708.x]
  - 9 **Schmitt-Gräff A**, Desmoulière A, Gabbiani G. Heterogeneity of myofibroblast phenotypic features: an example of fibroblastic cell plasticity. *Virchows Arch* 1994; **425**: 3-24 [PMID: 7921410]
  - 10 **Higashiyama R**, Nakao S, Shibusawa Y, Ishikawa O, Moro T, Mikami K, Fukumitsu H, Ueda Y, Minakawa K, Tabata Y, Bou-Gharios G, Inagaki Y. Differential contribution of dermal resident and bone marrow-derived cells to collagen production during wound healing and fibrogenesis in mice. *J Invest Dermatol* 2011; **131**: 529-536 [PMID: 20962852 DOI: 10.1038/jid.2010.314]
  - 11 **Guyot C**, Lepreux S, Combe C, Doudnikoff E, Bioulac-Sage P, Balabaud C, Desmoulière A. Hepatic fibrosis and cirrhosis: the (myo)fibroblastic cell subpopulations involved. *Int J Biochem Cell Biol* 2006; **38**: 135-151 [PMID: 16257564 DOI: 10.1016/j.biocel.2005.08.021]
  - 12 **Andrade ZA**, Guerret S, Fernandes AL. Myofibroblasts in schistosomal portal fibrosis of man. *Mem Inst Oswaldo Cruz* 1999; **94**: 87-93 [PMID: 10029917]
  - 13 **Kisseleva T**, Uchinami H, Feirt N, Quintana-Bustamante O, Segovia JC, Schwabe RF, Brenner DA. Bone marrow-derived fibrocytes participate in pathogenesis of liver fibrosis. *J Hepatol* 2006; **45**: 429-438 [PMID: 16846660 DOI: 10.1016/j.jhep.2006.04.014]
  - 14 **Forbes SJ**, Russo FP, Rey V, Burra P, Rugge M, Wright NA, Alison MR. A significant proportion of myofibroblasts are of bone marrow origin in human liver fibrosis. *Gastroenterology* 2004; **126**: 955-963 [PMID: 15057733 DOI: 10.1053/j.gastro.2004.02.025]
  - 15 **Russo FP**, Alison MR, Bigger BW, Amofah E, Florou A, Amin F, Bou-Gharios G, Jeffery R, Iredale JP, Forbes SJ. The bone marrow functionally contributes to liver fibrosis. *Gastroenterology* 2006; **130**: 1807-1821 [PMID: 16697743 DOI: 10.1053/j.gastro.2006.01.036]
  - 16 **Hinz B**, Phan SH, Thannickal VJ, Prunotto M, Desmoulière A, Varga J, De Wever O, Mareel M, Gabbiani G. Recent developments in myofibroblast biology: paradigms for connective tissue remodeling. *Am J Pathol* 2012; **180**: 1340-1355 [PMID: 22387320 DOI: 10.1016/j.ajpath.2012.02.004]
  - 17 **Bedossa P**, Poynard T. An algorithm for the grading of activity in chronic hepatitis C. The METAVIR Cooperative Study Group. *Hepatology* 1996; **24**: 289-293 [PMID: 8690394 DOI: 10.1002/hep.510240201]
  - 18 **Skalli O**, Vandekerckhove J, Gabbiani G. Actin-isoform pattern as a marker of normal or pathological smooth-muscle and fibroblastic tissues. *Differentiation* 1987; **33**: 232-238 [PMID: 3596085]
  - 19 **Lepreux S**, Bioulac-Sage P, Gabbiani G, Sapin V, Housset C, Rosenbaum J, Balabaud C, Desmoulière A. Cellular retinol-binding protein-1 expression in normal and fibrotic/cirrhotic human liver: different patterns of expression in hepatic stellate cells and (myo)fibroblast subpopulations. *J Hepatol* 2004; **40**: 774-780 [PMID: 15094224 DOI: 10.1016/j.jhep.2004.01.008]
  - 20 **van Eys GJ**, Niessen PM, Rensen SS. Smoothelin in vascular smooth muscle cells. *Trends Cardiovasc Med* 2007; **17**: 26-30 [PMID: 17210475 DOI: 10.1016/j.tcm.2006.11.001]
  - 21 **Council L**, Hameed O. Differential expression of immunohistochemical markers in bladder smooth muscle and myofibroblasts, and the potential utility of desmin, smoothelin, and vimentin in staging of bladder carcinoma. *Mod Pathol* 2009; **22**: 639-650 [PMID: 19252475 DOI: 10.1038/modpathol.2009.9]
  - 22 **Montani M**, Thiesler T, Kristiansen G. Smoothelin is a specific and robust marker for distinction of muscularis propria and muscularis mucosae in the gastrointestinal tract. *Histopathology* 2010; **57**: 244-249 [PMID: 20716166 DOI: 10.1111/j.1365-2559.2010.03618.x]
  - 23 **Maeng M**, Mertz H, Nielsen S, van Eys GJ, Rasmussen K, Espersen GT. Adventitial myofibroblasts play no major role in neointima formation after angioplasty. *Scand Cardiovasc J* 2003; **37**: 34-42 [PMID: 12745801]
  - 24 **Nam YH**, Lee SK, Sammut D, Davies DE, Howarth PH. Preliminary study of the cellular characteristics of primary bronchial fibroblasts in patients with asthma: expression of alpha-smooth muscle actin, fibronectin containing extra type III domain A, and smoothelin. *J Invest Allergol Clin Immunol* 2012; **22**: 20-27 [PMID: 22448450]
  - 25 **Yang L**, Scott PG, Dodd C, Medina A, Jiao H, Shankowsky HA, Ghahary A, Tredget EE. Identification of fibrocytes in postburn hypertrophic scar. *Wound Repair Regen* 2005; **13**: 398-404 [PMID: 16008729 DOI: 10.1111/j.1067-1927.2005.130407.x]
  - 26 **Bhunchet E**, Wake K. Role of mesenchymal cell populations in porcine serum-induced rat liver fibrosis. *Hepatology* 1992; **16**: 1452-1473 [PMID: 1446899 DOI: 10.1002/hep.1840160623]
  - 27 **Verhamme P**, Quarck R, Hao H, Knaepen M, Dymarkowski S, Bernar H, Van Cleemput J, Janssens S, Vermynen J, Gabbiani G, Kockx M, Holvoet P. Dietary cholesterol withdrawal reduces vascular inflammation and induces coronary plaque stabilization in miniature pigs. *Cardiovasc Res* 2002; **56**: 135-144 [PMID: 12237174 DOI: 10.1016/S0008-6363(02)00515-1]
  - 28 **Tharp DL**, Wamhoff BR, Turk JR, Bowles DK. Upregulation of intermediate-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel (IKCa1) mediates phenotypic modulation of coronary smooth muscle. *Am J Physiol Heart Circ Physiol* 2006; **291**: H2493-H2503 [PMID: 16798818 DOI: 10.1152/ajp-heart.01254.2005]
  - 29 **Otranto M**, Sarrazy V, Bonté F, Hinz B, Gabbiani G, Desmoulière A. The role of the myofibroblast in tumor stroma



- remodeling. *Cell Adh Migr* 2012; **6**: 203-219 [PMID: 22568985 DOI: 10.4161/cam.20377]
- 30 **Paner GP**, Shen SS, Lapetino S, Venkataraman G, Barkan GA, Quek ML, Ro JY, Amin MB. Diagnostic utility of antibody to smoothelin in the distinction of muscularis propria from muscularis mucosae of the urinary bladder: a potential ancillary tool in the pathologic staging of invasive urothelial carcinoma. *Am J Surg Pathol* 2009; **33**: 91-98 [PMID: 18936687 DOI: 10.1097/PAS.0b013e3181804727]
- 31 **Pinzani M**. Hepatic stellate (ITO) cells: expanding roles for a liver-specific pericyte. *J Hepatol* 1995; **22**: 700-706 [PMID: 7560865 DOI: 10.1016/0168-8278(95)80227-4]
- 32 **Wisse E**, Braet F, Luo D, De Zanger R, Jans D, Crabbé E, Vermoesen A. Structure and function of sinusoidal lining cells in the liver. *Toxicol Pathol* 1996; **24**: 100-111 [PMID: 8839287 DOI: 10.1177/019262339602400114]

**P- Reviewers:** Forte A, Grizzi F, Reyes VE **S- Editor:** Gou SX  
**L- Editor:** Stewart GJ **E- Editor:** Zhang DN





Published by **Baishideng Publishing Group Co., Limited**

Flat C, 23/F., Lucky Plaza,

315-321 Lockhart Road, Wan Chai, Hong Kong, China

Fax: +852-65557188

Telephone: +852-31779906

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

<http://www.wjgnet.com>



ISSN 1007-9327



9 771007 932045